



# Investigating the network properties of polymer matrixes for controlling a functionality of laccase-based amperometric biosensors

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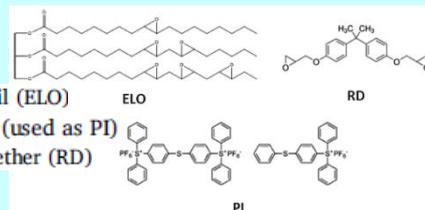
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<sup>4</sup>Kaunas University of Technology, 50254 Kaunas, Lithuania triarylsulfonium hexafluorophosphates (used as PI)

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## Motivation

The new direction in analytical biotechnology is the development of biosensors – bioanalytical devices that combine the best features of bioelements (selectivity) and physical transducers (high sensitivity and accuracy). Biosensors are complicated and effective device capable of fast detection and measuring wide spectrum in various applications in the field of healthcare, industrial process control, military application, environmental monitoring, agriculture and veterinary monitoring. Application of polymer as a holding matrix of immobilized enzyme is an innovative approach in a construction of the non-mediated enzyme-based biosensors of the third generation [1].

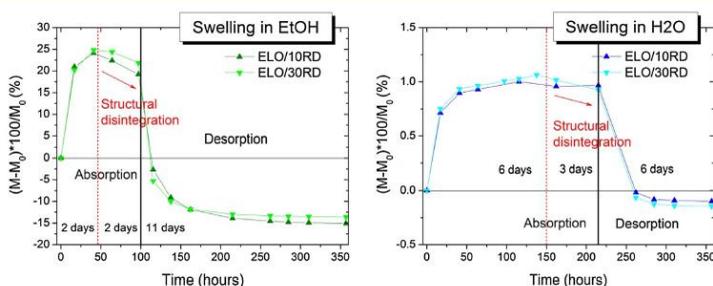


Fig. 2. The absorption and desorption of EtOH (left) and H<sub>2</sub>O (right) of the polymers ELO/10RD and ELO/30RD during 15 days.

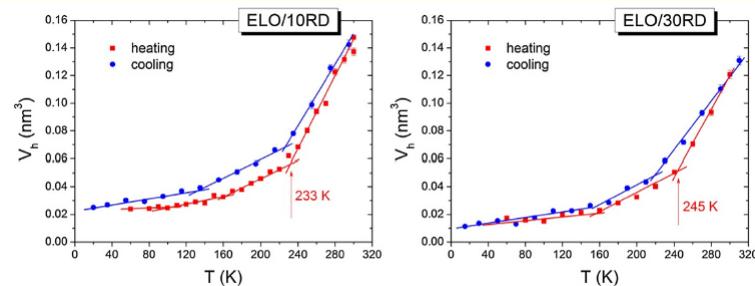


Fig. 3. Hole volume temperature dependences  $V_h(T)$  for the polymers ELO/10RD (left) and ELO/30RD (right) in the heating and cooling cycles.

Table 2

Hole volume  $V_h$  at glass transition temperature  $T_g$ , swellability  $S$  in EtOH, and slopes  $\alpha_{F1}$ ,  $\alpha_{F2}$  of the  $V_h(T)$  dependences in the regions below and above  $T_g$ , respectively, as well as their differences. Values for heating and cooling cycles are in the top and bottom part of the boxes, respectively.

Polymers	$V_h$ (nm <sup>3</sup> )	$T_g$ (K)	$S$ (%)	$\alpha_{F1}$ ( $10^{-4} K^{-1}$ )	$\alpha_{F2}$ ( $10^{-4} K^{-1}$ )	$\alpha_{F2} - \alpha_{F1}$ ( $10^{-4} K^{-1}$ )
ELO/10RD	$0.057 \pm 0.002$	233	24.09	$3.53 \pm 0.30$	$13.02 \pm 0.60$	$9.49 \pm 0.67$
	$0.068 \pm 0.002$			$3.31 \pm 0.32$	$11.16 \pm 0.55$	$7.85 \pm 0.64$
ELO/30RD	$0.051 \pm 0.002$	245	24.81	$3.47 \pm 0.33$	$12.42 \pm 0.64$	$8.95 \pm 0.72$
	$0.049 \pm 0.002$			$3.87 \pm 0.83$	$8.96 \pm 0.48$	$5.09 \pm 0.96$

Table 3

Biosensor response  $I_{max}$ , apparent Michaelis-Menten constant  $K_M^{app}$  toward ABTS as the substrate, the slope of the calibration curve  $B$ , the sensitivity of bioelectrodes (working surface area 7.35 mm<sup>2</sup>) constructed based on laccase immobilized by the polymers ELO/10RD and ELO/30RD, and the range of linearity of the constructed bioelectrodes to ABTS.

Polymers	$I_{max}$ ( $\mu A$ )	$K_M^{app}$ (mM)	$B$ ( $\mu A \cdot mM^{-1}$ )	Sensitivity ( $A \cdot M^{-1} \cdot m^{-2}$ )	Range of linearity (mM)
ELO/10RD	$4.9 \pm 0.19$	$0.36 \pm 0.03$	12.3	1.673	0.006–0.15
ELO/30RD	$1.25 \pm 0.17$	$0.11 \pm 0.04$	9.07	1.234	0.025–0.10

## Results and conclusion

In the present work, novel photocross-linked polymers are reported to be used in construction of laccase-based amperometric enzyme biosensors of the third generation for analysis of phenol derivatives [2]. It is found that the polymer ELO/10RD compared to the polymer ELO/30RD has: (i) the higher crosslink density, (ii) the larger free-volume holes, (iii) the lower concentration of free-volume holes, and (iv) the larger difference in the coefficients for the thermal expansion of free-volume holes in the regions below and above  $T_g$ . At the same time, the laccase-based amperometric biosensor constructed using the polymer ELO/10RD as a biosensor holding matrix shows the improved biosensor's parameters compared to ELO/30RD. Further research is required to prove this correlation for other polymers with more different crosslink density.

## References

- [1] T. Kavetsky et al., J. Appl. Polym. Sci., **134**, 45278 (2017).  
[2] T. Kavetsky et al., Eur. Polym. J., **115**, 391 (2019).

## Acknowledgments

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